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Investigating Amine Derivatives of Ambruticin VS-5 and VS-4

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A structure–activity relationship around the amine group of the ambruticin VS series has been developed for antifungal activity. It was shown that the amine can be alkylated through reductive amination without loss of potency. However, if it is converted into either an amide, carbamate, or urea, a significant loss of po-

tency is observed. Of the alkyl amines, small nonpolar groups are optimal for both potency and oral bioavailability. As a result of this study, one compound (KOS-2079) was taken into an animal efficacy model with success.

Introduction

The number of systemic fungal infections has increased rapidly over the past decade. This increase is principally due to the rise in immune-compromised patients, not only those with HIV/AIDS, but also patients undergoing treatment for cancer and organ transplant therapy.^[1] However, there are only a few drugs available to treat these infections, of which two classes are most widely used. The azoles, including fluconazole and itraconazole, have limited success against some fungal strains, especially Aspergillus spp. These compounds inhibit the enzyme 14 α -demethylase, preventing formation of ergesterol, an essential component of the fungal cytoplasmic membrane.^[2] Alternatively, amphotericin B, a polyene with a broad spectrum of activity but severe side-effects, is employed. Amphotericin is thought to associate with ergosterol-forming pores, which leads to leakage of K^+ ions and cell death.^[3] There remains a need for new and safe antifungal agents.

The ambruticins are a set of polyketide antifungal agents produced by the myxobacterium Sorangium cellulosum which vary in the substituent at the 5-position. The S series is characterized by a 5-hydroxy group, and was isolated in the 1970s by researchers at Warner-Lambert.^[4] In the 1990s a second series was isolated by researchers at GBF: the VS series, which posses an amine at the 5-position in various methylated states (Figure 1).^[5] The ambruticins have in vitro antifungal activities that are similar to amphotericin (with minimum inhibitory concentrations (MICs) in the μ g mL⁻¹ range).^[5,6] The mechanism of action of the ambruticins was first proposed by Knauth and

Figure 1. Ambruticin S (1; R=OH) and ambruticins VS-5 (2; R=NH₂), VS-4 $(3; R = NHMe)$, and VS-3 $(4; R = NMe₂)$.

Reichenbach to be similar to that of the phenylpyrroles, $[7]$ that is, through induction of the high osmolarity glycerol (HOG) signaling pathway.^[8] More recently, Vetcher et al. suggested that Hik1, a histidine kinase in the HOG pathway, is the likely target of ambruticin in S. cerevisiae.^[9]

There has been renewed interest in these molecules, resulting in several total syntheses of ambruticin S(1) being reported.^[10-13] Our group has investigated the structure-activity relationship (SAR) about the carboxylic acid in the VS series, $[14]$ building on the work of Warner-Lambert researchers on the S series.^[15, 16] These derivatives showed that the carboxylic acid is not essential for antifungal activity and that there was considerable tolerance for sterically small substituents. There has not been any discussion of the SAR in the amine region of the molecule; however, an ambruticin analogue, KOS-2079, was reported to be effective in two animal models. In the first, a mouse model of invasive aspergillosis,^[17] KOS-2079 significantly decreased pulmonary fungal burdens and improved survival over the vehicle control at 200 mg kg^{-1} day⁻¹. The second demonstrated that KOS-2079 prolonged the survival of mice infected with lethal doses of Coccidioides over vehicle-treated mice at both 20 and 50 mg kg⁻¹ day⁻¹.^[18] Herein we describe a series of amine derivatives that led to this compound being taken into these in vivo efficacy animal models.

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Results and Discussion

The amine functionality of the ambruticin VS series is an attractive site to effect transformations due to its orthogonal reactivity with the remainder of the molecule. It is also an ideal site to modulate the physical and pharmacokinetic properties of the ambruticins. In the natural series the amine appears in all three states of methylation (zero to two in 2–4), with little variation in antifungal activity (see Table 1), suggesting that there is no requirement for a hydrogen bond donor at this position. Thus, we set about derivatizing this site by commencing with both ambruticin VS-4 (3; monomethyl) and VS-5 (2; free amine), which were both readily accessible through fermentation of the producing organism.

The investigation began with reductive aminations using a range of aldehydes and ketones. The reaction proceeded well in the presence of the unprotected carboxylic acid at C1. In the VS-5 series it was difficult to obtain clean secondary amine product. Instead, reactions were allowed to proceed to the tertiary amine 5. As expected, the VS-4 series resulted in an Nmethyl-N-alkyl-substituted tertiary amine 6 (Scheme 1).

Scheme 1. Reductive aminations: a) R^1CHO , $Na(CN)BH_{3}$, CH_3OH , room temperature, 16 h; b) R²CHO or CH₃COCH₃, Na(CN)BH₃, CH₃OH, room temperature, 16 h.

The same methodology was used to synthesize a range of tertiary amines from both 2 and 3 to probe both the steric restrictions and the effects of introducing nonpolar, polar, and aromatic groups on the MICs required for antifungal activity against two Aspergillus strains (Table 1).^[19] The table shows that a range of N-alkyl amines can be tolerated. Steric bulk appears to be the determining factor in many cases. For example, comparison of VS-3 with the N,N-diethyl compound (4 versus 5a), and in the VS-4 series comparing N-ethyl, N-propyl, and Nbutyl (respectively 6a, 6b, 6c) shows a steady decline in potency. Branched or cyclic substituents are better tolerated than the equivalent straight-chain moieties (compare 6b with 6d, and $6c$ with $6e$ and $6f$). The N-(3-hydroxypropyl) compound 6 g suggests that the introduction of a polar group does not have a significant effect on potency. However, the introduction of an imidazole moiety (in 5 b and 6 h) results in a considerable decrease in potency. This may be due in part to the increase in steric bulk.

Table 1. Substituted amine derivatives of ambruticins VS-4 and VS-5 and

The reactivity of the amine allowed selective reaction with anhydrides, chloroformates, and isocyanates under standard reactions to form the corresponding amides (7 and 8), carbamates 9, and ureas 10 in either the VS-4 or VS-5 series without the need for protection (Scheme 2). In this way the effect of re-

Scheme 2. Amide, carbamate, and urea formation: a) $(R^2CO)_2O$, CH₃OH, room temperature, 20 h; b) CH₃OCOCl, iPr₂NEt, THF, room temperature, 20 h; c) R^2 NCO, THF, room temperature, 20 h. THF = tetrahydrofuran.

moving the free amine, which would be protonated under physiological conditions, could be assessed.

In the case of ambruticin VS-5 (2) the amine was converted into the azide 11 by reaction with trifluoromethanesulfonyl azide (Scheme 3).^[20, 21] These compounds were assessed for antifungal activity against our standard strains (Table 2). The results show that removing the amine leads to a significant decrease in potency in all cases. There is little to differentiate between the amides, carbamates, urea or azide. These compounds resemble the potency of ambruticin S(1), suggesting that the presence of an amine at C5 is required for the optimal potency.

Scheme 3. Azide formation: a) TfN₃, K₂CO₃, CuSO₄, H₂O/CH₂Cl₂, room temperature, 16 h. Tf $=$ trifluoromethanesulfonyl.

Based on the MIC results, five of the alkyl amine compounds were further investigated for their pharmacokinetic (PK) properties in mice. The molecules were chosen based on potency and also on range of substituent about the amine. Thus, there were two nonpolar compounds 5a and 6d displaying N-alkyl groups, a more polar compound 6 g having a 3-hydroxypropyl group, and two aromatic compounds 6h and 6i, the imidazolylmethyl and pyridylmethyl, respectively. By selecting compounds with a range of substituents we hoped to understand the effect that this portion of the molecule has on the PK properties.

All the molecules tested had high plasma exposures when administered intravenously and low volumes of distribution (V_{ss}) (Table 3). Many of the compounds have good oral bioavailability (%F), with the nonpolar side chains being optimal (5a, 6d versus 6g, 6h). All the molecules were well tolerated by mice. Based on the excellent potency and high oral bioavailability, compound 6d (given the number KOS-2079) was chosen for efficacy studies in two mouse models.^[17,18] Both studies

Table 3. Pharmacokinetic parameters for selected ambruticin analogues

showed that the molecule is able to prolong the life of the animals relative to vehicle.

Conclusions

In summary we have shown that the amine functionality of the VS series is vital for the increased potency of the VS series over the S series of ambruticin. If the amine is acylated the potency decreases to an MIC similar to that of ambruticin S. However, provided that the amine is retained it will tolerate a wide range of substitution. There is no requirement for a proton donor at this site and, in general, it appears that small nonpolar alkyl groups result in optimal potency and oral bioavailability. An example of this type, KOS-2079 (6 d) has been shown to prolong survival when given orally to mice in two lethal palmary infection models.

The ambruticins do appear to be an interesting scaffold on which to investigate a new range of antifungal agents. They possess an alternative mechanism of action to the widely used clinical agents, and they have high in vitro potency and excellent oral bioavailability. This study shows that there is room for tuning the properties in the VS series, and one of the molecules in this study has been shown to be efficacious in two animal models.^[17,18]

Experimental Section

Unless otherwise noted, ¹H and ¹³C NMR spectra were recorded in $CDCI₃$ at 300 K using a Bruker DRX 400 spectrometer. HRMS data were obtained by flow injection with manual peak-matching using an Applied Biosystems Mariner TOF spectrometer with a turbo-ion spray source. HPLC purification was performed using a Varian Metasil Basic reversed-phase column eluting with a gradient of acetonitrile in water with 0.1% acetic acid.

The care and husbandry of animals were in conformity with the institutional guidelines in compliance with national and international

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laws and policies. All animal work was conducted in an Association for Assessment and Accreditation of Laboratory Animal Care International (AAALAC, File #1205) accredited facility at Kosan.

General procedure for reductive aminations on ambruticins VS-5 and VS-4

The appropriate aldehyde or ketone (0.20 mmol, 2.0 equiv) was added to a solution of ambruticin VS-5 (2) or VS-4 (3) (0.10 mmol, 1 equiv) in CH₃OH (1.0 mL), followed by acetic acid (0.40 mmol, 4.0 equiv) and sodium cyanoborohydride (0.20 mmol, 2.0 equiv). The resulting solution was stirred at room temperature or 50° C until the starting ambruticin was consumed (typically 12 h). The reaction mixture was concentrated under reduced pressure and filtered through a C_{18} plug. The crude material was purified by HPLC.

5a; ¹H NMR (CD₃OD): δ = 3.31 (m, 2H, 2 x 1H of NCH₂), 3.03 (m, 2H, 2×1H of NCH₂), 1.34 ppm (t, 6H, $^{3}J_{H,H}$ =7.0 Hz, 2×CH₃); ¹³C NMR (CD₃OD): $\delta = 137.6$, 134.9, 134.8, 134.7, 129.4, 125.4, 123.9, 120.6, 81.2, 78.0, 73.5, 68.4, 63.4, 45.5, 43.7, 34.8, 30.3, 29.7, 29.3, 28.4, 25.1, 21.0, 20.3, 17.6, 11.9, 11.2, 9.6, 7.2 ppm; HRMS m/z 530 $[M+H]^+$ found 530.3842, C₃₂H₅₂NO₅ requires 530.3840 (Table 4).

5b; ¹H NMR (CD₃OD): $\delta = 7.92$ (s, 2H, 2×ArH), 7.10 (s, 2H, 2×ArH), 3.87 (d AB system, ${}^{3}J_{H,H}=14.0$ Hz, 2H, 2 \times 1H of NCH₂), 3.66 ppm (d AB system, ${}^{3}J_{H,H}=14.0$ Hz, 2H, 2 \times 1H of NCH₂); HRMS *m*/z 634 $[M+H]^+$ found 634.3966, C₃₆H₅₂N₅O₅ requires 634.3963 (Table 4).

6a; ¹H NMR (CD₃OD): $\delta = 3.33$ (m, 2H, NCH₂), 2.77 (s, 3H, NCH₃), 1.34 ppm (t, ${}^{3}J_{H,H}$ = 7.0 Hz, 3 H, CH₂CH₃); ¹³C NMR (CD₃OD): δ = 177.0, 137.8, 134.9, 134.8, 134.6, 129.4, 125.4, 123.8, 120.6, 81.1, 78.0 (2C), 73.1, 68.3, 65.4, 48.6, 43.3, 35.3, 34.8, 30.3, 29.7, 28.4, 25.1, 21.0, 20.2, 17.6, 11.9, 11.2, 9.1, 7.2 ppm; HRMS m/z 516 $[M+H]^+$ found 516.3701, C₃₁H₅₀NO₅ requires 516.3684 (Table 4).

6b; ¹H NMR δ = 3.33 (m, 1H, 1H of NCH₂), 2.89 (m, 1H, 1H of NCH₂), 2.63 (s, 3H, NCH₃), 1.72 (m, 2H, NCH₂CH₂CH₃), 0.92 ppm (t, $^3J_{\rm H,H}\!=\!7.0$ Hz, 3H, CH₃); ¹³C NMR $\delta\!=\!176.0$, 138.9, 135.5, 135.0 (2C), 129.5, 125.1, 123.8, 120.9, 81.4, 78.0, 77.8, 73.3, 69.2, 65.7, 55.6, 42.9, 36.7, 34.9, 30.5, 30.1, 28.9, 25.5, 21.4, 21.1, 18.9, 18.5, 12.9, 12.2, 11.2, 8.1 ppm; HRMS m/z 530 $[M+H]^+$ found 530.3838, $C_{32}H_{52}NO_5$ requires 530.3840 (Table 4).

6c; ¹H NMR δ = 2.94 (m, 2H, NCH₂), 2.61 (s, 3H, NCH₃), 1.65 (m, 2H, $CH_2CH_2CH_2$), 1.32 (m, 2H, CH_2CH_3), 0.91 ppm (t, ${}^3J_{H,H} = 7.5$ Hz, 3H, CH₃); ¹³C NMR δ = 176.0, 139.0, 135.5, 135.0 (2C), 129.5, 125.1, 123.7, 120.9, 81.5, 78.0, 77.8, 73.4, 69.2, 65.7, 54.0, 43.0, 36.7, 34.9, 30.5, 30.1, 29.0, 27.2, 25.5, 21.4, 21.1, 20.2, 18.9, 13.6, 12.9, 12.2, 8.1 ppm; HRMS m/z 544 $[M+H]^+$ found 544.3969, $C_{33}H_{54}NO_5$ requires 544.3997 (Table 4).

6d (KOS-2079); ¹H NMR δ = 3.34 (m, 1H, NCH), 2.48 (s, 3H, NCH₃), 1.31 (d, $\frac{3J_{H,H}}{6.5}$ Hz, 3H, 1 \times CH₃), 1.24 ppm (d, $\frac{3J_{H,H}}{6.5}$ Hz, 3H, $1 \times CH_3$; ¹³C NMR $\delta = 176.7$, 137.1, 135.1, 135.0, 134.9, 129.5, 125.3, 124.6, 120.8, 81.2, 77.9, 77.7, 73.3, 68.6, 63.8, 55.8, 43.1, 34.7, 32.3, 31.3, 30.5, 30.0, 28.7, 25.5, 21.1, 21.0, 18.8 (2C), 18.1, 12.9, 12.0, 8.1 ppm; HRMS m/z 530 $[M+H]^+$ found 530.3841, $C_{32}H_{52}NO_5$ requires 530.3840 (Table 4).

6e; ¹H NMR δ = 3.04 (m, 2H, NCH₂), 2.74 (s, 3H, NCH₃), 1.11 (m, 1H, 1H of cyclopropyl), 0.66 (m, 2H, 2H of cyclopropyl), 0.27 ppm (m, 2H, 2H of cyclopropyl); ¹³C NMR δ = 176.5, 139.8, 135.6, 135.0 (2C), 129.5, 125.0, 123.5, 120.9, 81.6, 78.0, 77.8, 73.6, 69.4, 65.0, 43.3, 34.9, 31.1, 30.5, 30.1, 29.1, 25.6, 21.5, 21.1, 18.9, 13.0, 12.2, 8.1, 7.0, 5.0, 4.2 ppm; HRMS m/z 542 $[M+H]^+$ found 542.3828, $C_{33}H_{52}NO_5$ requires 542.3840 (Table 5).

6 f; ¹H NMR δ = 2.67 (s, 3H, NCH₃), 2.29 (m, 1H, 1H of cyclopentyl), 2.09–2.02 (m, 2H, 2H of cyclopentyl), 1.78 (m, 2H, 2H of cyclopentyl), 1.55–1.48 (m, 2H, 2H of cyclopentyl), 1.29–1.24 ppm (m, 2H, 2H of cyclopentyl); ¹³C NMR δ = 176.0, 135.5, 135.1, 135.0, 129.5, 125.1, 123.6, 120.9, 81.5, 78.0, 77.8, 73.2, 68.9, 42.9, 34.9, 34.5, 30.5, 30.1, 29.1, 29.0, 28.9, 25.5, 23.5, 21.5, 21.1, 18.9, 13.0, 12.2, 8.1 ppm; HRMS m/z 556 $[M+H]$ ⁺ found 556.3981, $C_{34}H_{54}NO_5$ requires 556.3996 (Table 5).

6g; ¹H NMR (CD₃OD): δ = 3.86 (m, 2H, CH₂OH), 3.23 (m, 1H, 1H of NCH₂), 3.12 (m, 1H, 1H of NCH₂), 2.77 ppm (s, 3H, NCH₃); ¹³C NMR $(CD_3OD): \delta = 176.7, 137.5, 134.9, 134.8, 134.6, 129.4, 125.4, 124.0,$ 120.6, 81.1, 78.0 (2C), 73.3, 68.8, 66.4, 56.0, 54.9, 43.3, 36.6, 34.8, 30.3, 29.7, 29.2, 28.7, 28.4, 25.1, 20.9, 20.2, 17.6, 11.9, 11.2, 7.2 ppm; HRMS m/z 532 $[M+H]^+$ found 532.3638, $C_{31}H_{50}NO_6$ requires 532.3633 (Table 5).

6h; ¹H NMR (CD₃OD): δ = 7.82 (s, 1H, 1H of ArH), 7.30 (s, 1H, 1H of ArH), 4.22 (d AB system, ${}^{3}J_{H,H}=13.5$ Hz, 1 H, 1H of NCH₂), 4.10 (d AB system, ${}^{3}J_{H,H}$ = 13.5 Hz, 1H, 1H of NCH₂), 2.66 ppm (s, 3H, NCH₃); ¹³C NMR (CD₃OD): δ = 176.4, 137.4, 136.2, 134.9, 134.8, 134.6, 130.1, 129.4, 125.4, 124.1, 120.6, 119.6, 81.2, 78.0 (2C), 73.3, 68.9, 65.2, 49.9, 43.0, 35.3, 34.8, 30.3, 29.7, 29.2, 28.4, 27.7, 25.1, 20.9, 20.3, 17.7, 12.0, 11.2, 7.3 ppm; HRMS m/z 568 [M+H]⁺ found 568.3719, $C_{33}H_{50}N_3O_5$ requires 568.3745 (Table 5).

6i; ¹H NMR δ = 8.56 (s, 1H, ArH-2), 8.53 (d, $\frac{3}{4}$ _{H,H} = 4.0 Hz, 1H, ArH-6), 7.78 (d, ${}^{3}J_{H,H}$ = 6.5 Hz, 1H, ArH-4), 7.36 (t, ${}^{3}J_{H,H}$ = 5.5 Hz, ArH-5), 3.87 (m, 1 H, 1H of NCH₂), 3.68 (d AB system, $^{3}J_{\rm H,H}=$ 13.0 Hz, 1 H, 1H of NCH₂), 2.31 ppm (s, 3H, NCH₃); ¹³C NMR δ = 175.3, 174.2, 149.0, 147.8, 138.1, 137.8, 135.4, 135.0, 133.8, 129.6, 125.3, 124.3, 124.0, 120.9, 81.4, 78.0, 77.8, 73.0, 69.7, 65.4, 55.4, 41.1, 36.4, 34.9, 30.6, 30.1, 28.9, 28.8, 25.5, 21.4, 21.1, 18.9, 13.0, 12.1, 8.1 ppm; HRMS m/z 579 $[M+H]^+$ found 579.3827, $C_{35}H_{51}N_2O_5$ requires 579.3893 (Table 5).

6j; ¹H NMR δ = 7.80 (m, 3H, 3H of ArH), 7.74 (s, 1H, ArH-1), 7.56 (m, 3H, 3H of ArH), 4.10 (m, 1H, 1H of NCH₂), 3.84 (m, 1H, 1H of NCH₂), 2.33 ppm (s, 3H, NCH₃); ¹³C NMR δ = 175.6, 138.2, 135.4, 135.0 (2 C), 133.2, 132.9, 129.6, 128.4, 128.2, 127.8, 127.6 (2 C), 126.9, 126.2, 126.0, 125.3, 124.3, 120.981.5, 78.0, 77.8, 73.3, 69.6, 65.5, 58.2, 41.8, 36.4, 34.9, 30.6, 30.1, 28.9, 25.5, 21.4, 21.1, 18.9, 13.0, 12.2, 8.1 ppm; HRMS m/z 628 [M+H]⁺ found 628.3967, $C_{40}H_{54}NO_5$ requires 628.3996 (Table 5).

General procedure for amide formation with ambruticin VS-5 or VS-4

Acetic anhydride (1.00 mmol, 10 equiv) was added to a solution of ambruticin VS-5 (2) or VS-4 (3) (0.10 mmol, 1 equiv) in $CH₃OH$ (1.0 mL). The solution was stirred at room temperature for 20 h before concentrating under reduced pressure. The residue was dissolved in EtOAc (20 mL) and washed with H_2O (20 mL) and brine (20 mL) before drying ($Na₂SO₄$) and concentrating under reduced pressure. The crude product was purified by HPLC.

7a; ¹H NMR δ = 6.03 (brs, 1H, NH), 2.00 ppm (s, 3H, COCH₃); HRMS m/z 516 $[M+H]^+$ found 516.3339, $C_{30}H_{42}NO_6$ requires 516.3320 (Table 6).

8a; ¹H NMR $\delta = 2.91$ (s, 3H, NCH₃), 2.12 ppm (s, 3H, COCH₃); 13 C NMR δ = 174.2, 173.1, 138.3, 135.4, 135.0, 129.6, 125.2, 124.0, 120.9, 81.7, 78.0, 77.7, 72.4, 70.3, 55.4, 40.7, 34.9, 34.0, 30.6, 30.0, 29.0, 28.9, 25.5, 22.4, 21.4, 21.1, 18.9, 13.0, 12.2, 8.1 ppm; HRMS m/z 530 $[M+H]^+$ found 530.3500, $C_{31}H_{48}NO_6$ requires 530.3476 (Table 6).

7b; ¹H NMR δ = 5.82 (brs, 1H, NH), 2.25 (q, ${}^{3}J_{H,H}$ = 7.5 Hz, 2H, NCH₂), 1.15 ppm (t, ${}^{3}J_{H,H}$ = 7.5 Hz, 3H, CH₃); ¹³C NMR δ = 176.1, 173.4, 138.7, 135.4, 135.0, 129.6, 125.2, 123.7, 120.9, 81.4, 78.0, 77.8, 74.8, 71.8, 52.8, 40.0, 36.4, 34.9, 30.6, 30.0, 29.5, 29.0, 25.5, 21.4, 21.1, 18.9, 13.0, 12.2, 9.7, 8.1 ppm; HRMS m/z 530 $[M+H]^+$ found 530.3457, $C_{31}H_{48}NO_6$ requires 530.3476 (Table 6).

8b; ¹H NMR δ = 2.90 (s, 3H, NCH₃), 2.36 (m, 2H, OCH₂), 1.11 ppm (m, 3H, CH₃); ¹³C NMR δ = 176.1, 174.0, 138.2, 135.3, 134.9, 126.7, 125.3, 124.0, 120.9, 81.7, 78.0, 77.7, 72.2, 70.4, 55.5, 40.4, 34.9, 34.0, 30.6, 30.0, 29.7, 28.9, 27.4, 25.5, 21.4, 21.1, 18.9, 13.0, 12.1, 9.1, 8.1 ppm; HRMS m/z 566 [M+Na]⁺, 544 [M+H]⁺ found 544.3644, $C_{32}H_{50}NO_6$ requires 544.3633 (Table 6).

Procedure for carbamate formation with ambruticin VS-5 or VS-4

Diisopropylethylamine (0.30 mmol, 3.0 equiv) was added to a suspension of ambruticin VS-4 (3) (0.10 mmol, 1.0 equiv) in THF (1 mL) followed by methyl chloroformate (0.20 mmol, 2.0 equiv). The mixture was stirred at room temperature for 20 h before concentrating under reduced pressure. The residue was dissolved in EtOAc (20 mL), washed with HCl (0.1m, 20 mL) and brine (20 mL), dried (Na_2SO_4) , and concentrated under reduced pressure. The crude product was purified by HPLC to yield 9 (KOS-1972); ¹H NMR δ =

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3.70 (s, 3H, OCH₃), 2.81 ppm (s, 3H, NCH₃); HRMS m/z 568 $[M+Na]^+$ found 568.3227, C₃₁H₄₇NO₇Na requires 568.3248.

Procedure for the synthesis of the ureas

Alkyl isocyanate (0.50 mmol, 5.0 equiv) was added to a suspension of ambruticin VS-4 (3) (0.10 mmol, 1.0 equiv) in THF (1 mL). The mixture was stirred at 50 $^{\circ}$ C for 20 h before adding PS-TsNHNH₂ resin (Argonaut, CA, USA) and stirring at room temperature for 16 h. The mixture was diluted with $CH₃OH$ (10 mL) and filtered to remove the resin. The filtrate was concentrated under reduced pressure, and the residue was dissolved in EtOAc (20 mL). The solution was washed with HCl (0.1m, 20 mL) and brine (20 mL), dried $(Na₂SO₄)$, and concentrated under reduced pressure. The crude product was purified by HPLC.

10a; ¹H NMR δ = 5.85 (m, 1H, CHCH₂), 5.15 (d, ${}^{3}J_{H,H}$ = 17.0 Hz, 1H, trans-CH₂), 5.08 (d, ${}^{3}J_{H,H}=10.0$ Hz, 1H, cis-CH₂); 3.82 (s, 2H, NCH₂CH) 2.78 ppm (s, 3H, NCH₃); ¹³C NMR δ = 173.8, 159.9, 138.3, 135.4, 135.1, 134.9 (2C), 129.6, 125.2, 124.0, 120.9, 115.8, 81.7, 78.0, 77.8, 72.4, 70.7, 57.1, 43.5, 40.5, 34.9, 34.7, 30.5, 30.0, 28.9, 28.5, 25.5, 21.4, 21.1, 20.8, 18.9, 13.0, 12.2, 8.1 ppm; HRMS m/z 593 $[M+Na]^+$ found 593.3582, C₃₃H₅₀N₃O₆Na requires 593.3561 (Table 7).

10b; ¹H NMR δ = 7.32 (m 5 H, ArH), 5.14 (brs, 1 H, NH), 4.36 (m, 2 H, CH₂Ar), 2.73 ppm (s, 3H, NCH₃); ¹³C NMR δ = 175.0, 159.9, 139.1, 137.9, 135.3, 135.0, 134.9, 129.6, 128.6 (2C), 127.6 (2C), 127.2, 125.3, 124.3, 120.9, 81.5, 78.0, 77.7, 72.7, 70.6, 57.0, 45.0, 41.2, 34.9, 34.7, 30.5, 30.0, 28.9, 28.5, 25.5, 21.4, 21.1, 18.9, 13.0, 12.2, 8.1 ppm; HRMS m/z 643 $[M+Na]$ ⁺ found 643.3746, C₃₇H₅₂N₂O₆Na requires 643.3718 (Table 7).

Procedure for the synthesis 5-azidoambruticin 11

Potassium carbonate (0.56 mmol, 1.0 equiv) was added to a solution of ambruticin VS-5 (2) (0.56 mmol, 1.0 equiv) in H_2O (2.0 mL) and CH₃OH (4.0 mL), followed by TfN₃ (3.0 mL of a 0.37 m solution in CH_2Cl_2 , 1.11 mmol, 2.0 equiv). The mixture was made homogeneous by the addition of $CH₃OH$ (2 mL), and copper sulfate (0.006 mmol, 0.01 equiv) was added. The solution was stirred at room temperature for 16 h before concentrating under reduced pressure. The residue was loaded onto silica. Column chromatography (silica, $5 \rightarrow 10\%$ CH₃OH/CH₂Cl₂) yielded 11; ¹³C NMR $\delta = 174.1$, 140.3, 138.8, 135.1, 135.0, 129.6, 125.0, 123.0, 120.9, 81.4, 78.0, 77.9, 73.9, 71.6, 62.6, 39.9, 35.8, 35.0, 30.4, 30.1, 29.2, 25.6, 21.7, 21.1, 18.9, 13.0, 12.3, 8.2 ppm; HRMS m/z 501 [M+H]⁺ (Table 7).

General procedure for the antifungal activity assay

The NCCLS protocol was used.^[19] Compounds were dissolved in DMSO and prepared as a stock solution in RPMI media at $2 \times$ concentration. A stock solution of the spores for the filamentous fungi $(1-10\times 10^{4} \text{ cells} \text{ mL}^{-1})$ was made in RPMI media at 2x concentration. Aliquots of 100 µL of each solution were added to each well to a final volume of 200 mL. Drug-free and fungi-free controls were included. Plates were incubated at 37 \degree C for 48 h. To determine the MIC for filamentous fungi, the optical density (OD) of each well was measured at 405 nm with a spectrophotometer. After subtraction of the OD of the control not containing fungal spores, the percentage growth relative to the drug-free well was calculated. Relative OD values were grouped into five levels: $0 = 0-5\%$; $1 = 6-25\%$;

2=26-50%; $3 = 51-75$ %; $4 = 76-100$ %. The MICs reported refer to 50% inhibition.

General procedure for PK studies

Compounds were dissolved in ethanol at $20 \times$ above the targeted concentration. One part of the concentrate was diluted with 19 parts of a diluent consisting of 12% hydroxypropyl-ß-cyclodextrin in phosphate-buffered saline. Mice were dosed as indicated in Table 3 through either oral gavage or i.v. bolus injection through the tail vein. Plasma specimens were taken at various time points (up to 24 h post-dose). The plasma concentration of each compound was determined by LC–MS–MS using a Waters Alliance 2795 HPLC instrument coupled to a Waters QuattroMicro mass spectrometer. Pharmacokinetic parameters were calculated using Kinetica 4.1 (Thermo Electron).

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